

Functional polymorphism within *NUP210* encoding for nucleoporin GP210 is associated with the risk of endometriosis

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Objective: To investigate whether nucleoporin 210 (GP210, encoded by *NUP210* gene) is involved in endometriosis.

Design: Immunohistofluorescence analysis for assessing whether GP210 is expressed in endometrial tissues from patients and controls; genotyping and case-control study for assessing the association between rs354476 within *NUP210* and risk of endometriosis; in vitro luciferase assay for assessing the functional activity of rs354476.

Setting: University.

Patient(s): Histologically diagnosed cases (n = 175) of endometriosis: minimal or mild (stage I-II) in 48 cases (28%), moderate (stage III) in 69 cases (39%), and severe (stage IV) in 58 cases (33%). Controls (n = 557) were female blood donors collected at Meyer Hospital of Florence.

Intervention(s): None.

Main Outcome Measure(s): GP210 tissue expression; genotype distribution and risk of endometriosis; in vitro gene expression measurements.

Result(s): GP210 had positive nuclear immunohistofluorescence staining in endometrial glandular epithelium. Carriers of the variant allele were associated with increased risks: C/T, odds ratio (OR) 1.83, 95% confidence interval (CI) 1.04–3.21; T/T, OR 2.55, 95% CI 1.36–4.80. In vitro, luciferase assay showed that rs354476 is a bona fide target for hsa-miR-125b-5p.

Conclusion(s): Nucleoporin GP210 is involved in endometriosis. Rs354476 polymorphism affects the regulation of *NUP210* gene expression by altering the binding with hsa-miR-125b-5p, a microRNA already known as playing an important role for endometriosis. This provides the rationale for the observed increased risk of endometriosis in carriers of the variant allele. (Fertil Steril® 2019;112:343–52. ©2019 by American Society for Reproductive Medicine.)

El resumen está disponible en Español al final del artículo.

Key Words: Endometrium, glycoprotein-210, miRNA binding sites, *NUP210*, polymorphism

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Endometriosis is a common inflammatory disease, with endometrium-like tissues ectopically located outside the uterus. This condition is heterogeneous, with various manifestations that include

superficial and peritoneal lesions, endometriosis cysts in the ovaries (endometrioma, OMA), and nodules (>5 mm) in deep infiltrating endometriosis (DIE) (1, 2). The pathogenesis of endometriosis is unclear, and the research supports two main theories. In “retrograde menstruation,” viable endometrial cells invade other tissues (e.g., ovaries) through the fallopian tubes. The Müllerian-remnant theory hypothesizes that DIE represents adenomyosis originated in the

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retroperitoneum from embryonic rests of the Müllerian duct or extension of adenomyotic nodules arising in the myometrium. There is also an additional “iatrogenic theory” suggesting that previous pelvic surgery is a risk factor for disseminating endometrial cells outside the uterus (3). The predisposition to this condition likely has multifactorial bases (4, 5). Several risk factors have been identified, such as early age at menarche (<12 years) and a short menstrual cycle (<26 days), perhaps contributing to enhance the retrograde menstruation phenomenon. Exposure to polychlorinated biphenyl and dioxin is debated: these agents could play a role through the disruption of circulating hormone levels and/or dysregulation of the immune system. An inverse correlation between the risk of endometriosis and body mass index was also reported (6).

Pioneering studies observed that there is aggregation of endometriosis cases within families (7). In fact, the first-degree relatives of affected women showed an approximately six-fold higher risk than the first-degree relatives of unaffected women, suggesting an important role of the genetic background. This observation was corroborated by studies on twins that reported a heritability coefficient of approximately 50% (7). When the first genetic analyses were attempted by carrying out linkage analysis in families, the results showed weak but positive signals within the chromosomal regions 10q26 and 7p13-15, suggesting that the disease is multigenic (8). More recently, genome-wide association studies (GWAS), and meta-analyses helped to unravel part of this genetic complexity. These studies detected increased risk of endometriosis associated with specific alleles of common single nucleotide polymorphisms (SNPs). It is hypothesized that the associations could be driven by the genotyped SNP or by others in elevated linkage disequilibrium, because they could affect the function of the gene to which they belong. These studies are very helpful to spot specific genes and can contribute to generate hypotheses on the etiology of the disease.

A role of sex steroid hormones was suggested by positive associations between the risk of endometriosis and intronic variants within *ESR1* (rs1971256), *FSHB* (rs74485684), and *GREB1* (rs11674184) genes. The regulation of embryogenesis and cell fate was hypothesized given the positive signals found upstream of *HOXA10* (rs12700667), or within *WNT4* (rs12037376), *FN1* (rs1250241), and *ID4* (rs760794). Other signals were also suggestive for a role of inflammatory cytokines (*IL1A*, rs10167914), cell cycle regulators (*CDKN2B-AS1*, rs1537377; *ETAA1*, rs6546324), or angiogenesis (*KDR*, rs1903068) (9–11).

Overall, these genetic polymorphisms explain only a fraction (up to 5.19%) of the total variance, clearly indicating that more studies need to be undertaken. Recent evidences suggested that SNPs within microRNA (miRNA) binding sites (miRSNPs) could play a role. These SNPs weaken or strengthen the binding between messenger RNAs (mRNAs) and their targeting miRNAs (12), affecting protein translation or mRNA half-life. Thus, their effects on gene expression could provide a rational base for the observed associations between specific miRSNP genotypes and the risk of developing various human diseases, including (but not limited to)

cardiovascular diseases (13, 14), cancer (15–21), and, as said, endometriosis.

The MiRSNPs associated with endometriosis include [1] rs14647 within *WHSC1* (22), [2] rs3813486, rs1127473, and rs3211066 within *SLC22A23* (22), [3] rs7201 within *MMP2* (23), [4] rs334348 within *TGFBR1* (24), and [5] rs1434536 within *BMPRI1B* (25), affecting the binding sites of hsa-miR-99b, hsa-miR-125a-3p, hsa-miR-520g, hsa-miR-628-5p, and hsa-miR-125b-5p, respectively. In particular, this latter miRNA was found repeatedly involved in endometriosis. A previous study showed that hsa-miR-125b-5p was up-regulated (up to 10-fold) in serum of women affected by endometriosis as compared with healthy women, suggesting that it could be an independent diagnostic biomarker (26, 27). This miRNA was also associated with endometrial receptivity in hyper-stimulated women undergoing IVF-ET (28). Moreover, it was involved in endometrial dysfunctions as well as cancer (28–32). Furthermore, at least 19 of the more than 900 predicted targets for hsa-miR-125b-5p (Targetscan, freely available at <http://www.targetscan.org>) were involved in endometriosis, as reported in the scientific literature (Supplemental Table 1, available online). All these findings support the notion that hsa-miR-125b-5p is one of the most important miRNAs involved in endometriosis, and they strongly suggest that the identification of its targets could help in elucidating the pathogenic mechanisms.

In this context, the study of miRSNPs involving hsa-miR-125b-5p could constitute a useful strategy to this aim. A previous work carried out by our group showed that the expression of a fluorescent reporter gene chimerized with the 3'-UTR (untranslated region) of *NUP210* bearing the T-rs354476 was reduced when compared with the C-rs354476 counterpart. Moreover, in silico predictions suggested that the polymorphism could be a target site for hsa-miR-125b-5p. In summary, rs354476 within the 3'-UTR of *NUP210* is a strong candidate miRSNP for hsa-miR-125b-5p, suggesting that *NUP210* should be further investigated in the context of endometriosis (15). *NUP210* maps at 3p25.1 and encodes the nucleoporin GP210 (alias the nuclear pore glycoprotein-210). Nucleoporins are involved in the structural organization of the so-called nucleolar channel system. Described since 1961 (33), the nucleolar channel system consists of several layers of tubular membrane cisternae within the nuclei of cells constituting the normal endometrium glandular epithelium (EGE) (34). This organelle appears in the mid-secretory phase of the menstrual cycle when the endometrium is receptive to implantation of the fertilized egg (35, 36), and it can be observed prematurely when women undergo ovarian hyperstimulation (37). Its absence or its delayed development characterized several cases of unexplained primary infertility (38–40), and it is considered as a major hallmark of the postovulatory endometrium (41). Thus, it is not surprising to observe that nucleoporins were variably involved in several gynecologic disorders. For example, the endometrium of women affected by recurrent miscarriages showed over-expression of *NUP210* in a specific time window as compared with healthy controls (42). In summary, because rs354476 is a putative miRSNP of nucleoporin GP210 for miR-125b-5p, it could represent an ideal candidate

to be studied in relation to endometriosis. Thus, in the present work, we verified and showed for the first time that GP210 is expressed in the normal EGE. Then we explored the association of rs354476-*NUP210* with the risk of endometriosis, and we assayed whether rs354476 could be a bona fide miR SNP for miR-125b-5p.

MATERIALS AND METHODS

Patients

The study was carried out in consecutive women who attended the endoscopic surgical services of the Obstetrics and Gynecology Unit of the San Raffaele Scientific Institute and of the Department of Obstetrics and Gynaecology of the University of Siena between December 2009 and December 2010. All the patients underwent complete presurgery clinical examination before the diagnostic operative laparoscopy. Indications to laparoscopy included chronic pelvic pain, infertility, ovarian cysts, and myomas. All patients were regularly menstruating, without desire for pregnancy. None were taking medications except for sporadic nonsteroidal anti-inflammatory drugs. Patients with dysfunctional uterine bleeding, endometrial pathology, or autoimmune disease were excluded from the study. Three physicians highly skilled in the evaluation and treatment of endometriosis staged all patients according to the revised American Society for Reproductive Medicine classification (43). Endometriosis was documented in 175 women. Stage of the disease was found to be minimal or mild (stage I-II) in 48 cases (28%), moderate (stage III) in 69 cases (39%), and severe (stage IV) in 58 cases (33%). Laparoscopies were performed by experienced surgical teams, and diagnoses were always confirmed by standard histologic examination of biopsies from representative endometriotic lesions. The endometrial specimens presented here were withdrawn from consecutive women presenting at the clinical practice in the period December 2009 and December 2010. They were classified as proliferative or secretory according to the last menstrual period and confirmed by ultrasound scans using a transvaginal probe at 4.5–7.0 MHz and by histologic examination. Diagnosis of superficial peritoneal lesions of endometriosis was based on direct visualization when endometriotic implants presented as typical lesions. Diagnosis of ovarian endometrioma was defined by the occurrence of at least one ovarian cyst lined by endometriotic tissue. Diagnosis of DIE was defined when lesions infiltrated to a depth of at least 5 mm, beneath the peritoneal surface, in at least one of the subsequent sites: the bladder muscularis propria; the anterior rectovaginal pouch, posterior vaginal fornix, or retroperitoneal area between the anterior rectovaginal pouch and the posterior vaginal fornix; the uterosacral ligament; or the bowel muscularis propria (44).

Immunohistofluorescence

For immunohistofluorescence localizations of GP210 protein we analyzed healthy endometrial tissues from four volunteers (two women were in the proliferative, two in secretory phase), OMA lesions from four patients plus their endometrial eutopic counterparts (two patients were in the

proliferative phase, two in secretory phase), and DIE lesions from four patients with their eutopic endometrial counterparts (two patients were in the proliferative phase, two in secretory phase).

Tissues were fixed in 10% neutral buffered formalin at room temperature for 24 hours, paraffin-embedded, sectioned, and mounted on Superfrost Plus microscope slides (Fisher Scientific). After deparaffinization with xylene and dehydration with ethanol, sections were treated with PT Module buffer 2 (Thermo Scientific)–ethylenediaminetetraacetic acid, pH 8. Samples were then incubated in 5% bovine serum albumin in PBS (phosphate-buffered saline) for 30 minutes and finally incubated overnight at 4°C with the rabbit anti-NUP210 polyclonal antibody (Novus Biologicals) (1:50 in PBS–1% bovine serum albumin) or polyclonal rabbit IgG (Novus Biologicals) as negative control. Glass slides were then washed in PBS and incubated for 1 hour at room temperature with fluorescein isothiocyanate conjugate–labeled goat anti-rabbit IgG antibody (1:100) (Sigma). After washing in PBS, the sections were stained with 6-diamino-2-phenylindole for nuclei counterstaining in mounting medium (Santa Cruz Biotechnology) and then observed with a Leica DMB 6000 microscope. TIFF images were captured with a CFTR 6500 digital camera (Leica), and the levels of the intensities were quantified on the green channel with ImageJ software, version 1.52 (National Institutes of Health; <https://imagej.nih.gov/ij/>, 1997–2018.). The level of the background in our samples was evaluated by using the same isotype antibody at the same protein concentration as the primary antibody followed by the secondary antibody, as usual. The background was subtracted, and the net fluorescence was expressed as arbitrary units.

Case–Control Association Study

The polymorphism rs354476 was evaluated in a case–control association study. Controls were female blood donors collected at Meyer Hospital of Florence, with a minimal age of 18 years, whose details were published previously (45). According to the Helsinki declaration, both healthy and affected volunteers gave their written informed consent to participate in the study, and the study protocol was cleared by the local ethics committee (Azienda Ospedaliero-Universitaria Pisana, Comitato Etico Area Vasta Nord-Ovest, CEAVNO; Azienda Ospedaliera Universitaria Senese, Comitato Etico Area Vasta Sud_Est, CEASVE).

DNA Extraction and genotyping

Genomic DNA was isolated from peripheral blood using the Puregene Blood Kit (Gentra Systems). Quantitation was performed with a Qubit fluorimeter and its specific kit (Quant-iT dsDNA HS [Life Technologies]). Thirteen blood samples from patients did not yield DNA of quality for genotyping (<0.1 ng/μL), thus the genotypes are available for 162 patients and 557 controls. Genotyping of rs354476 was carried out using Pre-design TaqMan SNP Genotyping Assays, assay ID C_8755527_10 (Life Technologies). The genotyping of 15 samples (2%) was repeated blindly as quality

control. The results showed complete concordance between replicates.

In Silico Predictions

According to our previous work, hsa-miR-125b-5p had a differential binding with *NUP210*-3'-UTR according to the genotype of rs354476, whereas the closely related hsa-miR-125a-5p (belonging to the same family) could bind to *NUP210*-3'-UTR but is not affected by the SNP (10). The prediction was repeated here, by using "RNA-cofold" (freely available at <http://rna.tbi.univie.ac.at/cgi-bin/RNAcofold.cgi>) (46) to assess the binding free energies between *NUP210*-mRNA-C-allele or *NUP210*-mRNA-T-allele and either hsa-miR-125b-5p or hsa-miR-125a-5p.

Vectors Used and in Vitro Assays

We cloned the 3'-UTR regions of *NUP210* carrying alternatively C-rs354476 or T-rs354476 in the vector pmiRGLO. Thus, the luciferase from *Photinus pyralis* (Firefly luciferase) with the reporter gene Fluc was chimerized with the *NUP210*-3'-UTR. The vector expresses *Renilla reniformis* (Renilla luciferase), Rluc reporter as internal standard. Cells were transiently transfected with pmiRGLO chimeric construct and with the synthetic hsa-miR-125b-5p, or alternatively with the hsa-miR-125a-5p (Qiagen) or with "All Stars Negative Control siRNA," both used as negative controls (NC). Each experimental point was repeated six times, and each experiment was repeated three times. Forty-eight hours after transfection, cells were lysed, and the activity of Fluc and Rluc was measured, using a dual-luciferase reporter assay kit and a luminometer (Berthold Technologies). For both reporter genes, the ratio of the luminescence values was calculated as Fluc/Rluc luminescence ratio (FRLR) in cells treated with the miRNA of interest (FRLR-miRNA) or treated with the NC (FRLR-NC). The final measure of relative activity was expressed with the parameter R-NORM, calculated as the ratio FRLR-miRNA/FRLR-NC, and this parameter was used for the statistical analyses. All the technical details of cloning and luminescence measurements are reported in the [Supplemental Materials](#), available online.

Statistical Analyses

The statistical power of the case-control association study was calculated with the tool "PS—power and sample size calculation" (freely available <http://biostat.mc.vanderbilt.edu/PowerSampleSize>) (47), and it showed that the sample size of cases and controls was large enough to provide a power of 80% to detect an odd ratio (OR) of 1.86 and 92% to detect an OR of 2.04 in a dominant model (with type I error = 0.01).

To verify whether genotypes followed Hardy-Weinberg equilibrium (HWE), we applied the χ^2 test (1 degree of freedom) on the genotype counts of the whole sample set. We used multivariate logistic regression analysis (MLR) to examine the associations between the genotypes and the risk of cancer. The association analyses were based on the estimation of the ORs and their 95% confidence intervals

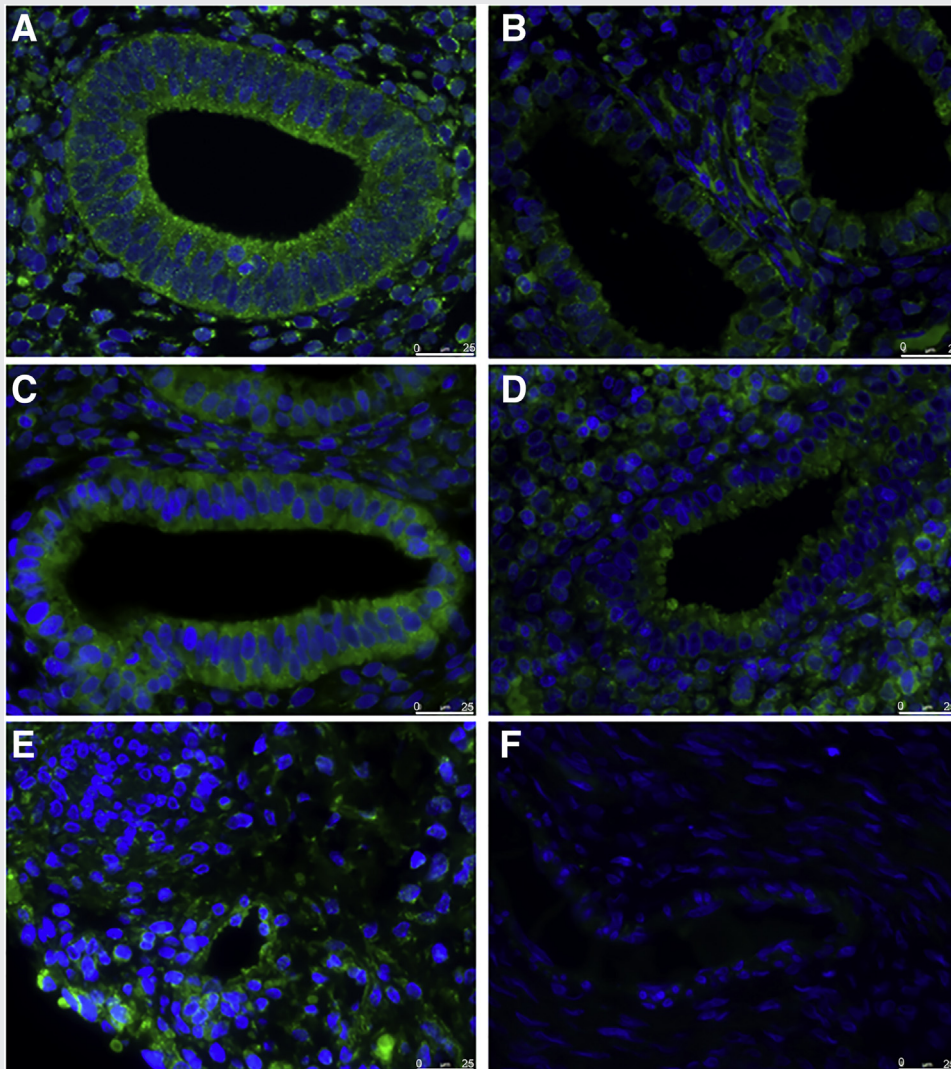
(CIs). In MLR, the ORs were adjusted for covariates as linear variables (OR_{adj}). Each genotype category was compared using the common homozygotes as reference category, and the *P*-value of the association (*P*_{ass}) was calculated separately for heterozygotes and homozygotes, in an additive model. Moreover, the most likely mode of inheritance was evaluated by performing an extended MAX test (48), which is based on multiplicity-adjusted *P* values for the Cochran-Armitage trend test statistics (49) of the dominant, additive, and recessive models. When the MAX test showed that the dominant or the recessive model was significant, then we performed the MLR analysis for that model. R-NORM was calculated by averaging the results obtained from three independent experiments, and each experiment was carried out using 12 independent replicates. R-NORM values were compared between genotypes using multifactor analysis of variance (MANOVA). All statistical tests were two-tailed and carried out using Statgraphics Centurion software (StatPoint).

RESULTS

According to previous results, *NUP210*-3'-UTR is a strong candidate target for hsa-miR-125b-5p. Here we evaluated further whether the gene could be a bona fide target for this miRNA by analyzing the data within the TCGA database (freely available at <https://portal.gdc.cancer.gov/>). We found that increased expression of hsa-miR-125b-5p correlates with reduced levels of *NUP210* mRNA in a series of 281 specimens from corpus uteri. This negative correlation (depicted in [Supplemental Fig. 1](#)) is highly statistically significant (*P* = .004), and it shows that the inter-individual variations of hsa-miR-125b-5p explain, alone, almost 3% ($r^2 = 2.58$) of the total inter-individual variability of *NUP210* mRNA. This result is in agreement with the fact that miRNAs could act as negative regulators of gene expression also by leading to reduced levels of their targeting messenger RNAs. Thus, the hypothesis that *NUP210* is a target for hsa-miR-125b-5p has been further corroborated by this analysis.

Then, we proceeded by assessing whether GP210 is actually expressed within EGE. We evaluated, by immunohisto-fluorescence staining, the expression of GP210 in healthy and eutopic endometrium, as well as in ectopic lesions. Representative pictures are reported in [Figure 1](#), and the corresponding fluorescence values (for the green channel) measured in the glandular epithelium and in the stroma are graphed in [Supplemental Figure 2](#). In the proliferative phase of the healthy endometrium, GP210 staining was markedly present in the EGE ([Fig. 1A](#)). The signal was positive within the cytoplasm of epithelial cells, mostly on the luminal side of the glands. Here we could measure an average of 112.5 units of fluorescence (SE = 6.39). Stromal cells, surrounding the glands, presented a poor expression, limited to small areas or spots around the nuclei, where we measured an average of 39 units (SE = 5.12, *P* < .001). In the secretory phase ([Fig. 1B](#)) the signal showed the same distribution but with an overall reduced intensity. In the cytoplasm of EGE cells the reduction was statistically significant (average 57.3 units, SE = 4.01, *P* < .001), whereas in the stroma the reduction was of a lesser

FIGURE 1



Representative images of immunohistofluorescence localization of GP210 (green) in healthy (A, B) and in eutopic endometrium from endometriotic patients (C, D), in OMA (E) and in DIE (F) ectopic lesions. Nuclei were counterstained with 6-diamino-2-phenylindole (blue). Bar = 25 μ m.

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extent (average 24.96 units, SE = 9.35, $P = .08$). Proliferative endometrium from patients suffering from endometriosis did not show specific differences as compared with EGE from healthy women. In fact, the pattern and the intensities of GP210 staining were remarkably similar in both the secretory (Fig. 1C) and proliferative (Fig. 1D) phases, with non-statistically significant differences when compared with their healthy counterparts (in the cytoplasm of EGE cells: average 92.6 units, SE = 5.6, and average 55.8 units, SE = 20.7, respectively; in the stroma: average 39.5 units, SE = 3.16, and average 31.8 units, SE = 10.4, respectively). When compared with healthy endometrium, OMA (Fig. 1E), showed a less organized EGE with a slightly reduced GP210 cytoplasmic fluorescence (average 76 units, SE = 14.1, $P < .001$) and similar intensity in the stroma (average 37.2 units, SE = 11.2, $P = .09$). The most obvious difference with the eutopic endometrium was found for peritoneal DIE, where poorly orga-

nized EGE structures could be detected, but lacking any cytoplasmic expression of GP210. The immunostaining of GP210 was low in the stroma (average 10.3 units, SE = 2.83) (Fig. 1F).

In silico predictions and our previous results suggested that rs354476, within the NUP210-3'-UTR, could affect the binding site for the hsa-miR-125b-5p. Here we replicated the calculations using RNA-cofold and confirmed that this latter miRNA had higher binding free energy for T (-10.84 kcal/mol) as compared with C (-10.03 kcal/mol) allele ($\Delta\Delta G = 0.81$ kcal/mol). At the same time, hsa-miR125a-5p is predicted to bind the NUP210-3'-UTR, but it shows similar binding free energies for both alleles (-13.32 kcal/mol for C- vs. -13.27 kcal/mol for T-allele), thus its use as internal control for the in vitro experiments is warranted.

The miRSNP rs354476 was then evaluated in relation to the risk of developing endometriosis in a case-control

TABLE 1

Case-control association study for rs354476.

Variable	Controls	Cases	OR ^a (95% CI)	P value
Age (y)	46.14 ± 12.08	32.53 ± 5.46		<10 ⁻⁴
Allele frequency rs354476-T	0.482 (537/1114)	0.514 (180/350)	1.34 (1.05–1.72)*	.0198*
Genotypes				
C/C	148	30	Ref	
C/T	281	84	1.83 (1.04–3.21)*	.035*
T/T	128	48	2.55 (1.36–4.80)*	.0036*
Best model	Additive	P-trend=.019*		

Note. Values for age are mean ± standard deviation and for allele frequency are rate (number). The difference of age means was compared with Student's t test. The adjusted ORs and their 95% CIs were calculated with multivariate logistic regression analysis. CI = confidence interval; OR = odds ratio.

*Statistically significant.

^a Adjusted for covariates.

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association study. In the present study cases were slightly younger than controls (32.5 ± 5.46 years [group average ± SD] and 46.14 ± 12.08 years, respectively), the genotype frequencies among controls followed Hardy–Weinberg equilibrium, and the allele frequencies were in agreement with those described for the same population, Tuscans (i.e. TSI, Tuscany) within dbSNP (freely available at <http://www.ncbi.nlm.nih.gov/SNP/> and in the database of 1000Genomes at <http://www.internationalgenome.org/>). After MLR, the heterozygotes (C/T) and homozygotes (T/T) showed a statistically significant association with the risk of endometriosis (OR_{adj} 1.83, 95% CI 1.04–3.21, *P* = .035 and OR_{adj} 2.55, 95% CI 1.36–4.80, *P* = .0036, respectively). Demography and results are reported in Table 1. When looking at the different clinical stages, increased risks were observed in all grading groups for TT genotype, with a highly statistical significance for patients at stage IV (Table 2). In this stage almost all volunteers (50 of 54) were carriers of the T-allele, and their risk was particularly increased (dominant model: OR_{adj} 6.58, 95% CI 1.94–22.30, *P* = .002). No differences in the average age groups were noticed according to the staging (Table 2).

To further understand the functional activity of the C/T variation we performed in vitro experiments by using chimeric vectors cotransfected with miRNAs. We tested hsa-miR-125b-5p for its differential binding with T or C alleles

and used hsa-miR-125a-5p as reference miRNA not having such an allelic-discriminating activity. Moreover, a random sequence miRNA, the known “All Stars Negative Control siRNA,” was also used as further negative control. Thus, we measured the normalized expression level (R-NORM) of the gene reporter in a dual luciferase assay in HCT116 cells after cotransfections with miRNAs and chimerized vectors. The results are represented in Figure 2. The activity is expressed as percent compared with the results obtained after the cotransfection of miRNA having the random sequence (referred to as 100%). We found that, as expected, hsa-miR-125b-5p and hsa-miR-125a-5p caused reduced expression of the reporter gene (MANOVA, *P* = 3.7 × 10⁻³ and *P* = 2 × 10⁻⁴, respectively). However, according to the in silico predictions, the two alleles responded differently to the different miRNAs (MANOVA with two-way interaction, *P* = .0173). In fact, after cotransfection with hsa-miR-125b-5p, the fluorescence of the C-allele was similar (97.3%) to that of the miRNA having random sequence, whereas that measured for the T-allele had a statistically significant reduction of almost 20%. Conversely, after cotransfection with hsa-miR-125a-5p, the fluorescence of the reporter gene chimerized with the 3'-UTR carrying the C-allele was reduced to a similar extent (approximately 17%) as that measured for the T-allele (approximately 16%).

TABLE 2

Characteristics of patients affected by endometriosis and association analysis (with multivariate logistic regression analysis) between genotypes of rs354476 and risk of endometriosis, stratified for grade of the disease.

Genotypes	Stages I–II (age: 32.5 ± 5.9 y)				Stage III (age: 32.0 ± 5.2 y)			Stage IV (age: 33.1 ± 5.5 y)		
	Controls (n)	n	OR ^a (95% CI)	P value	n	OR ^a (95% CI)	P value	n	OR ^a (95% CI)	P value
C/C	148	8	Ref.		18	Ref.		4	Ref.	
C/T	281	23	2.30 (0.86–6.13)	.10	26	0.80 (0.38–1.66)	.55	36	6.86 (1.99–23.64)*	.002*
T/T	128	12	2.91 (0.98–8.66)	.06	22	1.93 (0.88–4.21)	.10	14	5.90 (1.55–22.49)*	.009*
Total	557	43			66			54		

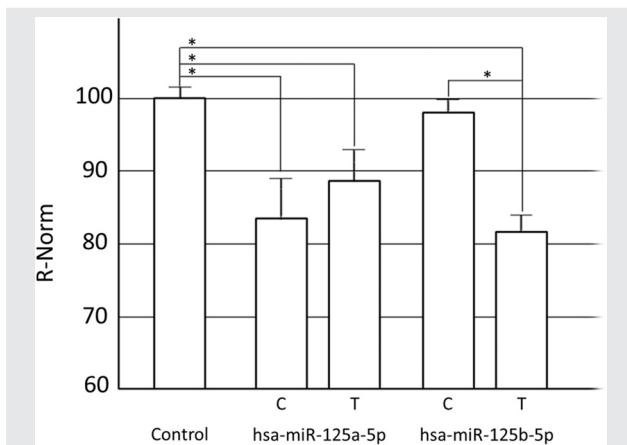
Note. Values for age are mean ± standard deviation. CI = confidence interval; OR = odds ratio.

*Statistically significant.

^a Adjusted for age.

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FIGURE 2



R-NORM of NCV compared with cells treated with a control miRNA (set at 100%). Each bar shows the average and SD of six independent replicates. Results are graphed according to the 3'-UTR genotype of the chimeric vector. Statistical analysis was performed with a multifactor analysis of variance model. Asterisk denotes a statistically significant difference between groups ($P < .05$). The HCT116 cell line showed a decreased expression of the reporter gene after treatment with both miR-125a and miR-125b-5p, in particular in the presence of allele T.

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DISCUSSION

The present study strongly suggests that *NUP210* mRNA is a bona fide target for hsa-miR-125b-5p, a key-miRNA involved in endometriosis, and that rs354476 affects their binding behaving as an actual miRSNP. These hypotheses are supported by previously published findings and by the inverse correlation between the expression of *NUP210* mRNA and hsa-miR-125b-5p in the TCGA dataset. Further support was provided by our in vitro assay carried out with the use of the synthetic miRNAs and chimeric constructs. This study also showed that GP210 is expressed within EGE, and this result is also corroborated by those reported within "The Human Protein Atlas Portal" (available at <http://www.proteinatlas.org>). Healthy EGE and eutopic endometriosis did not show particular differences with each other, whereas a reduced staining was noticed in the OMA, and a lack of staining was found in the peritoneal DIE. In the stroma the expression was always very poor.

In agreement with the hypotheses of a role for hsa-miR-125b and *NUP210* in endometriosis, the case-control association study showed that the miRSNP rs354476 is associated with increased risks, particularly strong for the more severe forms. We are aware that the association study could present some limitations. For example, cases and controls did not match for age. However, healthy controls were older than cases, and endometriosis is often underdiagnosed (with a 6.7-year mean latency from onset of symptoms to definitive diagnosis) (50). Thus, this fact reassured about potential occult endometrioses among controls at the corresponding age of patients. In any case, the results were adjusted for this covariate. Another limitation could have been repre-

sented by the sample size; however, the statistical power of the study was adequate (80%) to detect an OR of > 1.86 .

On the other hand, the study also has several strengths. First, a positive association was found in a genetic locus close to *NUP210* after a large meta-analysis of previous GWAS, suggesting that this genomic region could be a hot spot for endometriosis susceptibility. Moreover, a previous GWAS reported that rs6806012, an SNP in very close linkage disequilibrium with rs354476, was associated with the risk of endometriosis, confirming present findings (51). In addition, the allele frequencies measured in this study are corroborated by data from genomic databases for Tuscans. In fact, the series of controls analyzed here shows a T-allele frequency similar to that found in 1000Genomes for this population. Endometriosis patients show a higher allele frequency; therefore, if we pooled together Tuscans and present controls the statistical difference vs. endometriosis would be even stronger. Furthermore, the epidemiologic data are reinforced by the in vitro assay: the co-transfection of the reporter gene chimera with rs354476 T-allele, whereas the C-allele was not affected (the expression of the reporter gene was similar to that of the negative control). On the other hand, cotransfection with hsa-miR-125a-5p was able to reduce the expression of the reporter gene to a similar extent for both alleles. Thus, putting together previous and present results, we have reasons to believe that the association is unlikely to be a chance finding. In summary, we suggest that miRSNP rs354476 could affect the regulation of *NUP210* expression, thereby constituting a small but meaningful risk factor for this complex disease.

Mechanistically, one could speculate that the hsa-miR-125b-5p has a poor affinity for allele C- rs354476, causing increased expression of GP210. Conversely, hsa-miR-125b-5p has a higher affinity toward the corresponding T-allele, leading to a reduced expression of GP210. If this hypothesis were correct it would mean that GP210 is not only a marker of healthy EGE but, more importantly, it could be causally involved in the maintenance of its healthy status. Specific studies on the role of nucleoporins in the endometriosis are lacking. However, several works reported a relationship between endometrial carcinoma and *NUP88*. An increased expression of this protein was associated with premalignant endometrial lesions, early-stage endometrial cancer, and myometrial invasion of endometrial carcinoma (52, 53). Nucleoporins have been widely documented in the context of cell differentiation and embryogenesis. In 2010 a role for *NUP98* in the embryonic development of zebrafish was shown (54). In 2012 *NUP210* was clearly identified as playing a key role in determining cell fate during embryogenesis (55). Moreover, it was shown that the expression of nucleoporins is localized to specific embryonic anatomic structures, and specific patterns of expression during specific embryogenesis stages were shown in *Xenopus laevis* (56) and in mouse (57). Furthermore, a specific role of *NUP153* in the early stages of embryogenesis was shown in rabbit (58) and in cow (59). In particular, a role in the development of primordial germ cells was shown for *NUP50* (60). Finally, nucleoporins are

gaining increasing attention also for their role in the cellular physiology of the reproductive system in humans (61).

Thus, present data seem to fit better with the müllerian-remnant theory, suggesting that endometriosis could be generated by slight defects in the regulations of the cell fate in women, during embryogenesis. More studies are warranted to deep the analysis on nucleoporins in the context of endometriosis.

REFERENCES

- Reis FM, Petraglia F, Taylor RN. Endometriosis: hormone regulation and clinical consequences of chemotaxis and apoptosis. *Hum Reprod* 2013;4: 406–18.
- Vercellini P, Viganò P, Somigliana E, Fedele L. Endometriosis: pathogenesis and treatment. *Nat Rev Endocrinol* 2014;10:261–75.
- Barra F, Scala C, Biscaldi E, Vellone VG, Ceccaroni M, Terrone C, et al. Ureteral endometriosis: a systematic review of epidemiology, pathogenesis, diagnosis, treatment, risk of malignant transformation and fertility. *Hum Reprod Update* 2018;24:710–30.
- Hansen KA, Eyster KM. Genetics and genomics of endometriosis. *Clin Obstet Gynecol* 2010;53:403–12.
- Pagliardini L, Gentilini D, Viganò P, Panina-Bordignon P, Busacca M, Candiani M, et al. An Italian association study and meta-analysis with previous GWAS confirm WNT4, CDKN2BAS and FN1 as the first identified susceptibility loci for endometriosis. *J Med Genet* 2013;50:43–6.
- Zondervan KT, Becker CM, Koga K, Missmer SA, Taylor RN, Viganò P. Endometriosis. *Nat Rev Dis Primers* 2018;4:9.
- Giudice LC, Kao LC. Endometriosis. *Lancet* 2004;364:1789–99.
- Borghese B, Zondervan KT, Abrao MS, Chapron C, Vaiman D. Recent insights on the genetics and epigenetics of endometriosis. *Clin Genet* 2017;91:254–64.
- Krishnamoorthy K, Decherney AH. Genetics of endometriosis. *Clin Obstet Gynecol* 2017;60:531–8.
- Sapkota Y, Steinthorsdottir V, Morris AP, Fassbender A, Rahmioglu N, De Vivo I, et al. Meta-analysis identifies five novel loci associated with endometriosis highlighting key genes involved in hormone metabolism. *Nat Commun* 2017;24:15539.
- Painter JN, Anderson CA, Nyholt DR, Macgregor S, Lin J, Lee SH, et al. Genome-wide association study identifies a locus at 7p15.2 associated with endometriosis. *Nat Genet* 2011;43:51–4.
- Ma XP, Zhang T, Peng B, Yu L, Jiang de K. Association between microRNA polymorphisms and cancer risk based on the findings of 66 case-control studies. *PLoS One* 2013;208:e79584.
- Miller CL, Haas U, Diaz R, Leeper NJ, Kundu RK, Patlolla B, et al. Coronary heart disease-associated variation in TCF21 disrupts a miR-224 binding site and miRNA-mediated regulation. *PLoS Genet* 2014;10:e1004263.
- Contu R, Condorelli G. ATP6V0A1 polymorphism and microRNA-637: a pathogenetic role for microRNAs in essential hypertension at last? *Circ Cardiovasc Genet* 2011;4:337–8.
- Landi D, Gemignani F, Pardini B, Naccarati A, Garritano S, Vodicka P, et al. Identification of candidate genes carrying polymorphisms associated with the risk of colorectal cancer by analyzing the colorectal mutome and microRNAome. *Cancer* 2012;118:4670–80.
- Vymetalkova V, Pardini B, Rosa F, Jiraskova K, Di Gaetano C, Bendova P, et al. Polymorphisms in microRNA binding sites of mucin genes as predictors of clinical outcome in colorectal cancer patients. *Carcinogenesis* 2017;38: 28–39.
- Wang M, Du M, Ma L, Chu H, Lv Q, Ye D, et al. A functional variant in TP63 at 3q28 associated with bladder cancer risk by creating an miR-140-5p binding site. *Int J Cancer* 2016;139:65–74.
- Jiang W, Zhang W, Wu L, Liu L, Men Y, Wang J, et al. MicroRNA-related polymorphisms in PI3K/Akt/mTOR pathway genes are predictive of limited-disease small cell lung cancer treatment outcomes. *Biomed Res Int* 2017;2017:6501385.
- Stegeman S, Amankwah E, Klein K, O'Mara TA, Kim D, Lin HY, et al. A large-scale analysis of genetic variants within putative miRNA binding sites in prostate cancer. *Cancer Discov* 2015;5:368–79.
- Cao J, Luo C, Yan R, Peng R, Wang K, Wang P, et al. rs15869 at miRNA binding site in BRCA2 is associated with breast cancer susceptibility. *Med Oncol* 2016;33:135.
- Wang B, Yang H, Shen L, Wang J, Pu W, Chen Z, et al. Rs56288038 (C/G) in 3'UTR of IRF-1 regulated by MiR-502-5p promotes gastric cancer development. *Cell Physiol Biochem* 2016;40:391–9.
- Zhao ZZ, Croft L, Nyholt DR, Chapman B, Treloar SA, Hull ML, et al. Evaluation of polymorphisms in predicted target sites for micro RNAs differentially expressed in endometriosis. *Mol Hum Reprod* 2011;17:92–103.
- Tsai EM, Wang YS, Lin CS, Lin WY, Hsi E, Wu MT, et al. A microRNA-520 mirSNP at the MMP2 gene influences susceptibility to endometriosis in Chinese women. *J Hum Genet* 2013;58:202–9.
- Sepahi N, Kohan L, Jahromi AR, Daneshbod Y, Hoveidi EN. mir-126 rs4636297 and TGFβRI rs334348 functional gene variants are associated with susceptibility to endometriosis and its severity. *Gynecol Endocrinol* 2017;33:429–32.
- Chang CY, Chen Y, Lai MT, Chang HW, Cheng J, Chan C, et al. BMP1B up-regulation via a miRNA binding site variation defines endometriosis susceptibility and CA125 levels. *PLoS One* 2013;5:e80630.
- Cosar E, Mamillapalli R, Ersoy GS, Cho S, Seifer B, Taylor HS. Serum microRNAs as diagnostic markers of endometriosis: a comprehensive array-based analysis. *Fertil Steril* 2016;106:402–9.
- Ohlsson Teague EM, Van der Hoek KH, Van der Hoek MB, Perry N, Wagaarachchi P, Robertson SA, et al. MicroRNA-regulated pathways associated with endometriosis. *Mol Endocrinol* 2009;23:265–75.
- Chen C, Zhao Y, Yu Y, Li R, Qiao J. MiR-125b regulates endometrial receptivity by targeting MMP26 in women undergoing IVF-ET with elevated progesterone on HCG priming day. *Sci Rep* 2016;6:25302.
- Xia X, Wang J, Liu Y, Yue M. Lower cystic fibrosis transmembrane conductance regulator (CFTR) promotes the proliferation and migration of endometrial carcinoma. *Med Sci Monit* 2017;23:966–74.
- Shang C, Lu YM, Meng LR. MicroRNA-125b down-regulation mediates endometrial cancer invasion by targeting ERBB2. *Med Sci Monit* 2012;18: 149–55.
- Chang KK, Liu LB, Jin LP, Meng YH, Shao J, Wang Y, et al. NME1 suppression of endometrial stromal cells promotes angiogenesis in the endometriotic milieu via stimulating the secretion of IL-8 and VEGF. *Int J Clin Exp Pathol* 2013;6:2030–8.
- Dong F, Zhang Y, Xia F, Yang Y, Xiong S, Jin L, et al. Genome-wide miRNA profiling of villus and decidua of recurrent spontaneous abortion patients. *Reproduction* 2014;148:33–41.
- Dubrausky V, Pohlmann G. [The ultra-structure of the corpus endometrium during the cycle]. *Arch Gynakol* 1961;196:180–99.
- Spornitz UM. The functional morphology of the human endometrium and decidua. *Adv Anat Embryol Cell Biol* 1992;124:1–99.
- Clyman MJ. A new structure observed in the nucleolus of the human endometrial epithelial cell. *Am J Obstet Gynecol* 1963;15:430–2.
- Gordon M. Cyclic changes in the fine structure of the epithelial cells of human. *Int Rev Cytol* 1975;42:127–72.
- Zapantis G, Szymga MJ, Rybak EA, Meier UT. Premature formation of nucleolar channel systems indicates advanced endometrial maturation following controlled ovarian hyperstimulation. *Hum Reprod* 2013;28: 3292–300.
- Kohorn EI, Rice SI, Hemperly S, Gordon M. The relation of the structure of progesterational steroids to nucleolar differentiation in human endometrium. *J Clin Endocrinol Metab* 1972;34:257–64.
- Gore BZ, Gordon M. Fine structure of epithelial cell of secretory endometrium in unexplained primary infertility. *Fertil Steril* 1974;25:103–7.
- Dockery P, Pritchard K, Warren MA, Li TC, Cooke ID. Changes in nuclear morphology in the human endometrial glandular epithelium in women with unexplained infertility. *Hum Reprod* 1996;11:2251–6.
- Guffanti E, Kittur N, Brodt ZN, Polotsky AJ, Kuokkanen SM, Heller DS, et al. Nuclear pore complex proteins mark the implantation window in human Endometrium. *J Cell Sci* 2008;121:2037–45.

42. Lédée N, Munaut C, Aubert J, Sérazin V, Rahmati M, Chaouat G, et al. Specific and extensive endometrial deregulation is present before conception in IVF/ICSI repeated implantation failures (IF) or recurrent miscarriages. *J Pathol* 2011;225:554–64.
43. Revised American Society for Reproductive Medicine classification of endometriosis. *Fertil Steril* 1997;67:817–21.
44. Chapron C, Fauconnier A, Vieira M, Barakat H, Dousset B, Pansini V. Anatomical distribution of deeply infiltrating endometriosis: surgical implications and proposition for a classification. *Hum Reprod* 2003;18:157–61.
45. Cipollini M, Pastor S, Gemignani F, Castell J, Garritano S, Bonotti A, et al. TPO genetic variants and risk of differentiated thyroid carcinoma in two European populations. *Int J Cancer* 2013;133:2843–51.
46. Lorenz R, Bernhart SH, Siederdisen CH, Tafer H, Flamm C, Stadler PF, et al. ViennaRNA package 2.0. *Algorithms Mol Biol* 2011;6:26.
47. Dupont WD, Plummer WD. Power and samples size calculations. A review and computer. *Control Clin Trials* 1990;11:116–28.
48. Hothorn LA, Hothorn T. Order-restricted scores test for the evaluation of population-based case-control studies when the genetic model is unknown. *Biomed J* 2009;51:659–69.
49. Armitage P. Test for linear trends in proportions and frequencies. *Biometrics* 1955;11:375–86.
50. Burney RO, Giudice LC. Pathogenesis and pathophysiology of endometriosis. *Fertil Steril* 1999;98:511–9.
51. Adachi S, Tajima A, Quan J, Haino K, Yoshihara K, Masuzaki H, et al. Meta-analysis of genome-wide association scans for genetic susceptibility to endometriosis in Japanese population. *J Hum Genet* 2010;55:816–21.
52. Li Y, Zhang X, Ge J, Liu X, Xu S, Zhu Z, et al. Can Nup88 expression be associated with atypical endometrial hyperplasia and endometrial cancer? A preliminary study. *Pathol Res Pract* 2016;212:274–8.
53. Schneider J, Martínez-Arribas F, Torrejón R. Nup88 expression is associated with myometrial invasion in endometrial carcinoma. *Int J Gynecol Cancer* 2010;5:804–8.
54. Fung TK, Chung MI, Liang R, Leung AY. Role of a novel zebrafish nup98 during embryonic development. *Exp Hematol* 2010;38:1014–21.
55. D'Angelo MA, Gomez-Cavazos JS, Mei A, Lackner DH, Hetzer MW. A change in nuclear pore complex composition regulates cell differentiation. *Dev Cell* 2012;22:446–58.
56. Reza N, Khokha MK, Del Viso F. Nucleoporin gene expression in *Xenopus tropicalis* embryonic development. *Int J Dev Biol* 2016;60:181–8.
57. Lupu F, Alves A, Anderson K, Doye V, Lacy E. Nuclear pore composition regulates neural stem/progenitor cell differentiation in the mouse embryo. *Dev Cell* 2008;14:831–42.
58. Popken J, Schmid VJ, Strauss A, Guengoer T, Wolf E, Zakhartchenko V. Stage-dependent remodeling of the nuclear envelope and lamina during rabbit early embryonic development. *J Reprod Dev* 2016;62:127–35.
59. Popken J, Graf A, Krebs S, Blum H, Schmid VJ, Strauss A, et al. Remodeling of the nuclear envelope and lamina during bovine preimplantation development and its functional implications. *PLoS One* 2015;10:e0124619.
60. Park E, Lee B, Clurman BE, Lee K. NUP50 is necessary for the survival of primordial germ cells in mouse embryos. *Reproduction* 2016;151:51–8.
61. Preston CC, Storm EC, Leonard RJ, Faustino RS. Emerging roles for nucleoporins in reproductive cellular physiology. *Can J Physiol Pharmacol* 2018;2:1–8.

El polimorfismo funcional en NUP210 codificando la nucleoporina GP210 se asocia con riesgo de endometriosis.

Objetivo: Investigar si la nucleoporina 210 (GP210, codificada por el gen NUP210) está involucrada en la endometriosis.

Diseño: Análisis de inmunohistoquímica para evaluar si GP210 se expresa en tejidos endometriales de pacientes y controles; genotipado y estudio de casos y controles para evaluar la asociación entre rs354476 dentro de NUP210 y el riesgo de endometriosis; ensayo de luciferasa in vitro para evaluar la actividad funcional de rs354476.

Lugar: Universidad.

Paciente (s): casos diagnosticados histológicamente (n=175) de endometriosis: mínima o leve (estadio I-II) en 48 casos (28%), moderada (estadio III) en 69 casos (39%), y grave (estadio IV) en 58 casos (33%). Los controles (n=557) fueron mujeres donantes de sangre recolectados en el Hospital Meyer de Florencia.

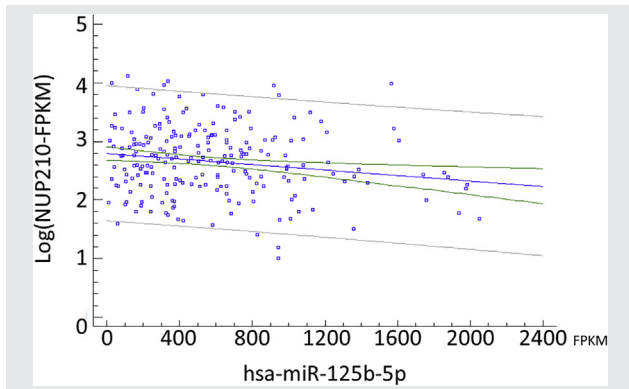
Intervención (es): Ninguna.

Principales medidas de resultado: Expresión tisular GP210; Distribución del genotipo y riesgo de endometriosis; medidas de expresión génica in vitro.

Resultados: GP210 tuvo tinción de inmunohistoquímica nuclear positiva en el epitelio glandular endometrial. Portadores de la variante en el alelo se asoció con mayores riesgos: C / T, odds ratio (OR) 1,83; intervalo de confianza (IC) del 95% 1,04–3,21; T / T; OR 2,55; IC del 95% 1,36–4,80. In vitro, el ensayo de luciferasa mostró que rs354476 es una diana adecuada para hsa-miR-125b-5p.

Conclusión: La nucleoporina GP210 está involucrada en la endometriosis. El polimorfismo Rs354476 afecta la regulación de la expresión génica de NUP210 al alterar la unión con hsa-miR-125b-5p, un microRNA ya conocido que juega un papel importante en la endometriosis. Esto justifica el aumento observado del riesgo de endometriosis en portadores del alelo variante.

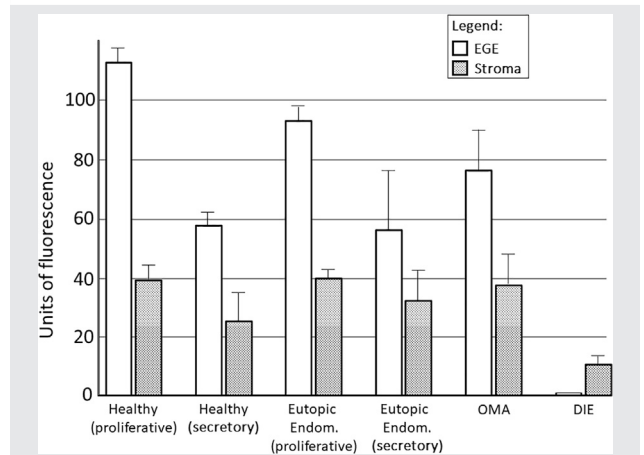
SUPPLEMENTAL FIGURE 1



Regression analysis between the expression of hsa-miR-125b-5p and *NUP210* mRNA according to the TCGA dataset (available at <https://portal.gdc.cancer.gov/>) after total RNA sequencing (with a next-generation sequencing approach) and expressed as FPKM (fragments per kilobase million). The analysis is performed on 281 samples from corpus uteri.

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SUPPLEMENTAL FIGURE 2



Graphic showing the measurements obtained with ImageJ on the TIFF images from the immunohistochemistry analysis. White bars are the values (in arbitrary units on the green channel) taken from EGE; dotted bars are the values taken from the surrounding stroma. The samples analyzed are reported in the bar chart and, in order, represent healthy endometrium in the proliferative phase (n = 2), healthy endometrium in the secretory phase (n = 2), eutopic endometrium in the proliferative phase from patients with OMA (n = 2) or DIE (n = 2), eutopic endometrium in the secretory phase from patients with OMA (n = 2) or DIE (n = 2), and the corresponding lesions OMA (n = 4) and DIE (n = 4).

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